

User Guide

Exo-spin™ mini-HD columns

Exosome Purification Kit

For cell culture medium, saliva, urine
and blood plasma/serum

Cat EX05



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Exo-spin™ mini-HD columns Exosome Purification Kit

Product components

EX05-5 Exo-spin™ mini-HD columns kit (5 columns)

- 5 x Exo-spin™ mini-HD columns
- 1 x User Guide

EX05-20 Exo-spin™ mini-HD columns kit (20 columns)

- 20 x Exo-spin™ mini-HD columns
- 1 x User Guide

EX05-100 Exo-spin™ mini-HD columns kit (100 columns)

- 100 x Exo-spin™ mini-HD columns
- 1 x User Guide

For all kits, large volume (15 ml or 50 ml) centrifuge tubes, PBS, and 1.5 ml microcentrifuge collection tubes are not supplied.

General exosome isolation information

A. Notes on cell culture

Fetal bovine serum (FBS) contains a large number of exosomes. Exosome-free FBS should be used in cell culture experiments, which can be obtained commercially. Alternatively, Vivaspin® 20 100kDa MWCO Polyethersulfone (GE Healthcare) or Amicon® Ultra-15 Centrifugal Filter Unit (Millipore) can be used to efficiently remove exosomes from FBS diluted 1:1 with PBS.

The number of exosomes that are obtained from a cell culture sample will vary depending on a variety of factors. These include the specific cell line, the length of time the cells are exposed to the medium, and cell density. Cancer cell lines may produce higher numbers of exosomes than non-transformed cell lines.

B. Notes on blood samples collection

Sample collection and handling prior to purification may have a significant impact on the quality of purified exosomes (Witwer *et al.*, 2013). Sera can contain many platelet-derived exosomes released after clot formation. Use of heparin-based anticoagulants is discouraged because of possible effects on downstream applications (e.g. PCR).

Platelet-derived exosomes may be released from platelets by the physical forces associated with the blood sampling procedure. Standardization of sampling site, needle gauge (wider may be better), and other variables is recommended. To ensure the sample is not contaminated by skin fibroblasts, it has been suggested that the first few milliliters of drawn blood should be discarded.

Collected blood should be handled gently and processed rapidly (within 30 minutes of drawing).

C. Proteomic analysis

Precipitants can interfere with mass spectrometry analysis and so precipitation should not be used when purifying exosomes if mass spectrometry is to be performed. In such cases, an alternative concentration method should be used instead of precipitation prior to using the Exo-spin™ columns. To maximize the numbers of exosomes that can be purified from cell culture media, devices such as the CELLline Classic bioreactor flask (Sigma) can increase the concentration of exosomes in media by up to 8-fold.

Product information

Exo-spin™ technology combines Precipitation and Size Exclusion Chromatography (SEC), making it superior to techniques that rely solely on precipitation which result in co-purification of large amounts of non-exosomal proteins and other material as well as carry over of the precipitant. However, EX05 kit only contains SEC columns as the precipitation step is not required for some applications (e.g. mass spectrometry downstream application and/or ≤150 µl blood samples).

Isolated exosomes may be used in a variety of downstream applications including DNA and RNA studies, as well as in functional *in vitro* and *in vivo* exosome assays.

This kit has been developed to process different sample types:

- 1 to 75 ml starting volume per column for cell culture medium, saliva, urine, and other low-protein biological fluids
- ≤150 µl starting volume per column for blood samples (sera and plasma)

Protocol for purification of intact exosomes using Exo-spin™ mini-HD columns

Supplied Exo-spin™ columns are pre-equilibrated with ultra-pure water containing 20% ethanol. The column matrix should be re-equilibrated with PBS prior to use.

A maximum sample volume of 75 ml may be used per column. For blood samples, the maximum sample volume is $\leq 150 \mu\text{l}$ per column. For larger sample volumes, use multiple columns per sample. **Please note that purchasing additional Exo-spin™ Buffer (cat EX06-30 (30 ml) or EX06-250 (250 ml)) is required for processing the aforementioned maximum volume of starting material for non-blood samples.**

All centrifugation steps can be performed at room temperature or 4°C unless otherwise specified.

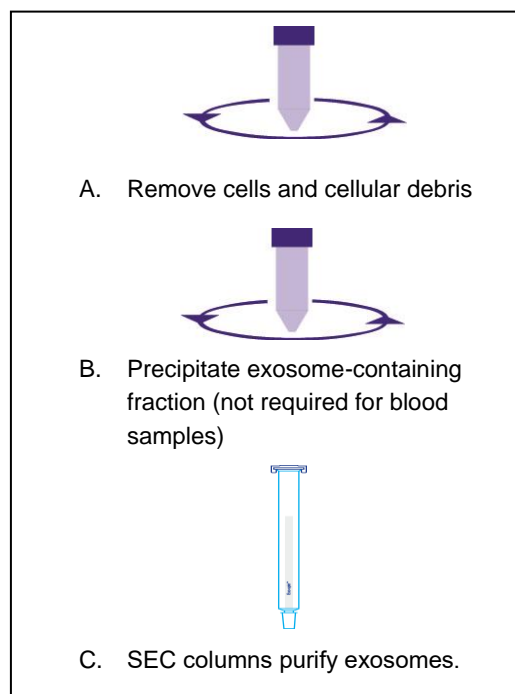


Figure. Protocol overview.

A. Prepare starting sample

Cell culture media, urine, saliva or blood sera

1. Transfer 1 – 75 ml of starting sample or 100 – 150 μ l of blood sera to a microcentrifuge (not supplied with kit) tube and spin at 300 \times g for 10 minutes to remove cells.
2. Transfer supernatant to a new microcentrifuge tube and spin at 16,000 \times g for 30 minutes to remove any remaining cell debris.

Blood plasma

Adapted from Welton *et al.*, 2015.

1. Centrifuge blood plasma at 6000 \times g for 10 minutes.
2. Filter the resulting platelet-free plasma through a 0.22 μ m syringe filter. Separate sample into 100 or 150 μ l aliquot.

B. Precipitate exosome-containing fraction (not required for blood sera/plasma, proceed directly to Part C)

3. Transfer supernatant to a new microcentrifuge tube and add Exo-spin™ Buffer in a 2:1 ratio (for example, add 25 ml of Exo-spin™ Buffer to 50 ml supernatant).
4. Mix well by inverting the tube and incubate at 4°C for at least 1 hour.
Alternatively, the sample may be incubated overnight at 4°C. This may generate a small increase in exosome yield.
5. Centrifuge the mixture at 16,000 \times g for 1 hour.
6. Carefully aspirate and discard the supernatant.
Do not allow the sample to dry as this may cause damage to exosomes.
7. Resuspend the exosome-containing pellet in 100 or 150 μ l of PBS. If the pellet does not readily resuspend, reduce the amount of starting material.

C. Exo-spin™ mini-HD column preparation

8. Prepare the Exo-spin™ mini-HD column prior to application of your sample.
 - a. Remove the cap and discard the preservative buffer from the top of the column. Remove the outlet plug. To prevent drying of the column bed, proceed to the next step immediately.
 - b. Equilibrate the column by sequentially adding 3 \times 2.5 ml of PBS and allow the column to drain under gravity. **Do not centrifuge.**

D. Purification of exosomes – high resolution fractionation

9. When the column has been equilibrated, fit the column onto a clamp-stand, with a 1.5 ml microcentrifuge tube underneath the column for high resolution fraction collection.
10. Carefully apply 100 or 150 μ l of the exosome-containing sample to the top of the column (filtered blood plasma from step 2 or resuspended exosome-containing pellet in PBS from step 7). Collect the fraction in the 1.5 ml microcentrifuge tube by gravity. Once the sample has entered the column, add 50 μ l of PBS and continue collecting first fraction for a total of 200 μ l. Label the tube as Fraction 1.

11. Place a new 1.5 ml microcentrifuge tube underneath the column. Apply 200 μ l of PBS to the column and collect the fraction by gravity. Label the tube as Fraction 2.
12. Repeat this process with PBS a further 22 times, until 24 fractions of 200 μ l each have been collected in total.

The vast majority of exosomes will elute in fractions 6 and 7. These fractions can be pooled to obtain a 400 μ l fraction which contains highly pure exosomes. If used with the maximum capacity volume (150 μ l) and a slightly higher yield is desired, fraction 8 can also be pooled to obtain a 600 μ l fraction. Be aware that this can lead to lower purity levels.

As an alternative to steps 9 – 12, a quick collection protocol can be performed:

9. When the column has been equilibrated, carefully apply 100 or 150 μ l of exosome-containing sample to the top of the column (filtered blood samples from step 2 or resuspended exosome-containing pellet in PBS from step 7). Place the column into a waste tube and allow the column to drain under gravity.
10. Apply 900 μ l of PBS to the top of the column if sample volume is 100 μ l or 850 μ l of PBS if sample volume added is 150 μ l. Allow the PBS to drain under gravity into a waste tube and discard.
11. Place a new 1.5 ml microcentrifuge collection tube. Add a further 400 – 600 μ l of PBS and allow the PBS to drain under gravity to collect the exosome-containing eluate.
For 100 μ l sample volume use 400 μ l of PBS. For 150 μ l sample volume, use 400 μ l to maximize exosome purity or 600 μ l to maximize exosome yield.

Iterative loading, flushing and storage

For samples larger than 150 μ l, iterative loading allows sample volumes up to 750 μ l to be loaded. Simply flush the columns with PBS after purification of the first 150 μ l of sample to remove free protein from the column and allow additional sample or new samples to be purified on the same column

- After step 12 (Section D) above, add a further 4 ml of PBS. This removes all free protein fractions from previous loadings (as determined by Bradford assay).
- The column may then be loaded with additional (or a new) sample by repeating the steps above.
- Note that flow rates may reduce if the column is re-used
- If storing the column for future use, use 20% ethanol to flush instead of PBS

Storage

Upon receipt, store Exo-spin™ mini-HD columns at 4°C.

Correctly stored components are stable for at least 3 months following purchase.

Related products

Related products	Product description	Product code
Exosome detection	Exosome antigen antibodies	EX201, EX202, EX204, EX203
	TRIFic™ detection assay	EX101, EX102, EX103
Nanoparticle Tracking Analysis (NTA) size profiling service	ZetaView NTA Particle Analysis Service	ZV-1 and ZV-12

TRIFic™ detection assay

The TRIFic™ exosome assay is similar to an ELISA, however, there are some significant differences. Unlike an ELISA, there is no enzymatic reaction. Rather, the target is directly detected with a Europium label. TRIFic™ exosome assays deliver clear, consistent, and quantitative data from purified or unpurified samples, including direct measurement of exosomes from plasma in a convenient 96-well format. TRIFic™ exosome assays are available for widely-used markers of exosomes, the tetraspanin proteins CD9, CD63 and CD81.

enables ExoFLARE™ to be monitored for extensive periods to allow tracking of dye movement.

NTA size profiling service

Exosome characterization service for analysis of particle size and particle concentration using the ZetaView instrument from Particle Metrix.

Troubleshooting

My sample does not elute from the column.

- Ensure that the outlet plug has been removed from the base of the column

My sample contains a lower amount of exosomes than expected.

- Ensure that the column does not dry out during the procedure.
- Adhere to the volumes indicated for sample addition to the column. If the sample volume is too small, the exosomes will be retained within the column.
- Ensure that precipitation of the exosome-containing pellet is performed for at least 1 hour at 4°C, and that the Exo-spin™ Buffer has been properly mixed with the sample.
- Exosome yield is dependent on a variety of factors, particularly the type of biological fluid used as starting material. If media is used, the amount of exosomes present will vary widely depending on the cell line and the length of exposure (conditioning) of cells to the media.

My sample has no measurable exosomes.

- This is most likely caused by complete drying out of the column causing loss of functionality. Ensure the columns are kept hydrated at all times.

I do not have a high-speed centrifuge.

- Increase the time of centrifugation by calculating the ratio of the recommended speed to the speed of your centrifuge. For example, if the protocol recommends to spin at 16,000 x *g* for 30 minutes, for a centrifuge with a maximum speed of 9,500 x *g*: $16000/9500=1.68$ and $1.68*30 \text{ mins} = 50.4 \text{ minutes}$.

References

- Witwer *et al.*, Journal of Extracellular Vesicles 2: 10.3402/jev.v2i0.20360 (2013).
- Welton *et al.*, Journal of Extracellular Vesicles 4: 10.3402/jev.v4.27269 (2015).

Purchaser Notification

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Cell Guidance Systems' reagents and services enable control, manipulation and monitoring of the cell, both *in vitro* and *in vivo*

Growth Factors

- Recombinant
- Sustained Release

Exosomes

- Purification
- Detection
- Tracking
- NTA Service

Small Molecules

Cell Counting Reagent

Matrix Proteins

Cell Culture Media

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- Photostable
- *In Vitro* Blastocyst Culture
- ETS-embryo Culture
- Custom Manufacturing Service

Gene Knock-Up System

Cytogenetics Analysis



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